

Design and Development of an Artificial Intelligence-Assisted Portable Molecular Diagnostic Platform for Rapid Detection of Plant Diseases and Precision Agriculture

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ARTICLE INFO

Article history:

Received 16 Aug 2026
Accepted 27 Aug 2026
Available online 31 Aug 2026

Keywords:

Robotics Process Automation (RPA), Artificial Intelligence (AI), Machine Learning (ML).

ABSTRACT

The rapid emergence of plant diseases, climate variability, and increasing global food demand have created an urgent need for intelligent, rapid, and field-deployable diagnostic technologies to support sustainable agriculture. Conventional plant disease detection methods, including visual inspection and laboratory-based molecular analysis, are often time-consuming, expensive, infrastructure-dependent, and unsuitable for real-time field applications. Although Polymerase Chain Reaction (PCR) and Quantitative Real-Time PCR (qPCR) provide highly sensitive and accurate pathogen detection, their dependence on centralized laboratories limits their practical application in precision agriculture. This study presents an Artificial Intelligence-Assisted Portable Molecular Diagnostic Platform for rapid and on-site detection of plant diseases. The proposed platform integrates automated DNA extraction, portable qPCR, a modular microfluidic biosensor cartridge, fluorescence-based detection, embedded electronics, wireless communication, cloud computing, and Artificial Intelligence within a compact portable architecture. The system combines molecular diagnostic data with biosensor signals and environmental information to support automated pathogen identification, disease classification, severity assessment, and intelligent agricultural decision-making. The proposed Artificial Intelligence framework incorporates Machine Learning and Deep Learning algorithms, including Random Forest, XGBoost, Support Vector Machine, Convolutional Neural Network, and Transformer-based approaches. Explainable Artificial Intelligence techniques are incorporated to improve the interpretability of diagnostic predictions. Experimental validation involves comparison of biosensor outputs with qPCR measurements using sensitivity, specificity, accuracy, detection limit, repeatability, and other statistical performance indicators. The results presented in the research demonstrate that the integrated platform provides rapid, reliable, and field-deployable molecular diagnosis while maintaining strong agreement with laboratory-based qPCR analysis. The integration of biosensors, Artificial Intelligence, wireless communication, and cloud-based analytics provides a comprehensive technological framework for precision agriculture, sustainable crop protection, and real-time disease surveillance.

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Introduction:

Plant diseases are among the major factors affecting agricultural productivity, food security, and sustainable crop production [24], [79]. Early and accurate identification of plant pathogens is essential for implementing timely disease management strategies and minimizing crop losses. Conventional disease diagnosis commonly depends on visual inspection, laboratory-based molecular testing, and expert interpretation. However, visual symptoms may appear only after significant pathogen development, while laboratory-based molecular techniques require specialized

instruments, trained personnel, controlled laboratory conditions, and transportation of samples.

Molecular diagnostic techniques such as Polymerase Chain Reaction (PCR) and Quantitative Real-Time PCR (qPCR) have significantly improved the sensitivity and specificity of plant pathogen detection [12], [17]. However, the requirement for centralized laboratory infrastructure limits their use in remote agricultural environments. Therefore, there is a growing requirement for portable molecular diagnostic systems capable of providing rapid and reliable results directly at the agricultural field.

Recent developments in biosensor technology, microfluidics, embedded electronics, Artificial Intelligence, Internet of Things (IoT), and cloud computing provide opportunities to overcome these limitations [13], [19], [76], [78], [84]. Portable biosensors can convert molecular interactions into measurable electrical or optical signals, while Artificial Intelligence can analyse molecular and environmental data to support automated disease classification and prediction.

The proposed research addresses these limitations by integrating automated magnetic bead-based DNA extraction, portable qPCR, fluorescence-based biosensing, embedded Artificial Intelligence, wireless communication, cloud computing, and Explainable Artificial Intelligence into a single portable molecular diagnostic platform. The integrated architecture is designed to provide rapid pathogen detection and intelligent agricultural decision support without complete dependence on centralized laboratories.

Research Gap

The literature review identified several limitations in existing plant disease diagnostic technologies. Many conventional molecular diagnostic systems remain dependent on centralized laboratories and manual sample preparation [12], [15], [17]. Portable qPCR systems improve field accessibility but may still require external DNA extraction and separate interpretation systems.

Another important limitation is the fragmented integration of molecular diagnostics, biosensors, embedded electronics, Artificial Intelligence, and cloud computing. Existing technologies generally focus on individual components rather than providing a unified sample-to-result platform [1], [15], [21].

Furthermore, Explainable Artificial Intelligence remains insufficiently implemented in portable agricultural molecular diagnostics [11], [67]. Existing AI systems may provide accurate predictions but often provide limited information regarding the biological and environmental factors responsible for those predictions.

Aim of the Study

The primary aim of this research is to design and develop an Artificial Intelligence-Assisted Portable Molecular Diagnostic Platform capable of rapid, accurate, and field-deployable detection of plant diseases for precision agriculture and sustainable crop management.

Objectives

The major objectives of the study are:

- To investigate the limitations of conventional plant disease diagnostic methods.
- To develop a compact and field-deployable molecular diagnostic device.
- To integrate automated DNA extraction.
- To integrate portable qPCR for pathogen-specific DNA detection.
- To develop a modular microfluidic biosensor cartridge.
- To develop AI-based pathogen classification and disease prediction.
- To implement Explainable Artificial Intelligence.

- To integrate wireless communication, cloud computing, and mobile applications.
- To evaluate sensitivity, specificity, accuracy, repeatability, reproducibility, detection limit, and response time.
- To compare the proposed platform with conventional PCR/qPCR-based laboratory diagnosis.

Related Work:

Molecular Diagnosis of Plant Disease

Molecular diagnostics provide reliable identification of plant pathogens through detection of pathogen-specific nucleic acids. PCR and qPCR are widely used because of their high sensitivity, specificity, and quantitative capability [12], [17]. However, conventional systems require laboratory infrastructure and trained personnel.

Portable Molecular Diagnostics

Portable molecular diagnostic technologies have emerged as alternatives to centralized laboratory systems [1], [15], [21]. These devices combine molecular biology, microfluidics, biosensors, embedded electronics, and wireless communication into compact platforms. Their primary advantage is the ability to perform molecular analysis near the point of sample collection.

Biosensor Technology

Biosensors combine a biological recognition element with a physicochemical transducer to convert biological interactions into measurable signals [20], [22], [45], [46]. Electrochemical, optical, paper-based, and microfluidic biosensors are particularly suitable for portable diagnostics because of their compact size, rapid response, and compatibility with embedded systems.

Microfluidic biosensors can integrate sample preparation, DNA extraction, amplification, pathogen detection, and signal analysis within compact Lab-on-a-Chip architectures [14], [19]. Their advantages include reduced reagent consumption, rapid processing, automation, and reduced contamination risk.

Artificial Intelligence in Molecular Diagnostics

Artificial Intelligence provides computational techniques for analysing complex molecular and environmental datasets. Machine Learning algorithms such as Random Forest, XGBoost, Support Vector Machine, and Deep Learning algorithms can identify patterns within molecular signals, qPCR amplification data, fluorescence measurements, and environmental parameters [10], [56], [58], [52].

The proposed system uses Artificial Intelligence to perform pathogen identification, disease classification, disease severity estimation, outbreak prediction, and agricultural decision support [6], [8], [87].

Research Gap

Existing technologies generally provide only selected components of the complete diagnostic process. The proposed platform addresses this gap by integrating automated DNA extraction, portable qPCR, biosensor detection, Edge AI, cloud computing, wireless

communication, and Explainable AI within a single portable architecture.

Proposed Ai-Assisted Portable Molecular Diagnostic Platform:

Overall Architecture

The proposed platform consists of interconnected modules including:

- Automated DNA extraction module
- Portable qPCR module
- Microfluidic biosensor cartridge
- Optical detection system
- Electrochemical sensing system
- Embedded processing unit
- Edge AI module
- Wireless communication module
- Cloud database
- Mobile application
- Decision-support dashboard

The overall architecture enables a sample-to-result diagnostic workflow in which biological samples are processed, pathogen-specific DNA is amplified, biosensor signals are detected, and Artificial Intelligence generates diagnostic and agricultural recommendations.

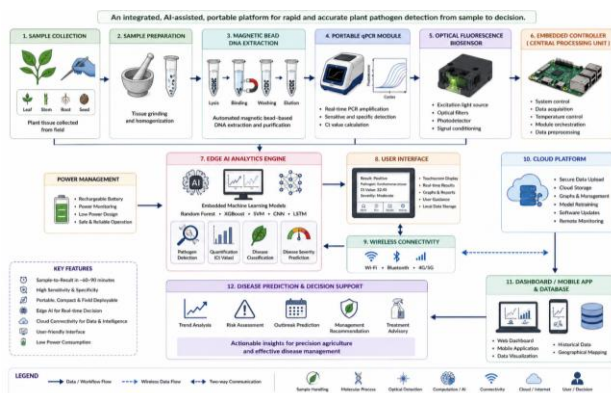


Fig 1_Overall Architecture of the Proposed AI-Assisted Portable Molecular Diagnostic Platform

Automated DNA Extraction

Automated DNA extraction is designed to isolate high-quality nucleic acids from plant and environmental samples. Magnetic bead-based extraction is incorporated to reduce manual handling and contamination risk. The extracted DNA is subsequently transferred to the molecular amplification and biosensor analysis stages.

Portable qPCR Module

The portable qPCR module provides pathogen-specific DNA amplification and quantitative measurement. Fluorescence is monitored during amplification and Cycle Threshold (Ct) values are obtained for quantitative interpretation.

The qPCR system serves as the laboratory reference method for validating the performance of the portable biosensor.

Microfluidic Biosensor

The microfluidic cartridge provides a compact platform for molecular recognition and sensing. Pathogen-specific DNA

probes interact with complementary target sequences, producing measurable optical and electrochemical signals. Gold nanoparticles and graphene-based sensing materials are incorporated to enhance signal response and molecular detection capability [3], [4], [26], [48].

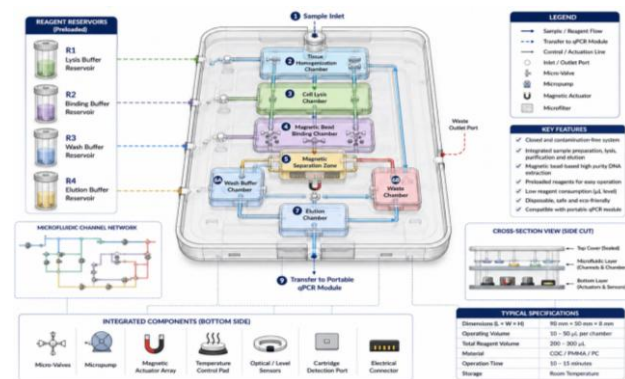


Fig 2_Architecture of the Proposed Microfluidic Biosensor

Artificial Intelligence Module

The AI module processes molecular, biosensor, and environmental data. The proposed framework integrates Random Forest, XGBoost, CNN, and Transformer-based models [56], [58], [51], [64], [65].

The model receives:

- DNA quality parameters
- qPCR Ct values
- Fluorescence intensity
- Biosensor signals
- Environmental parameters
- Weather information
- Soil characteristics
- Crop information
- Satellite-derived vegetation indices

The dataset is divided into training, validation, and testing subsets using a 70:15:15 ratio. Model performance is evaluated using Accuracy, Precision, Recall, Specificity, Sensitivity, F1-Score, ROC-AUC, MAE, RMSE, and confusion matrix analysis.

Edge AI and Cloud Integration

Edge AI performs immediate analysis directly on the portable device, reducing dependence on continuous internet connectivity. When connectivity is available, summarized diagnostic information is synchronized with cloud infrastructure [7], [76].

Cloud computing supports long-term data storage, disease surveillance, geographical mapping, model retraining, and centralized agricultural monitoring [84], [85], [81].

Experimental Methodology:

Sample Collection

Plant and environmental samples were collected from healthy and diseased plants representing different crops, environmental conditions, and disease stages. Samples included leaves, roots, stems, seeds, and soil-associated material. Sterile equipment and appropriate sample identification procedures were used to minimize contamination.

Leaf samples were collected from healthy and visibly infected plants and preserved under controlled conditions until molecular analysis.

DNA Extraction

Collected samples were processed using the proposed DNA extraction workflow. The objective was to obtain high-quality DNA suitable for qPCR and biosensor analysis.

qPCR Validation

qPCR was performed as the reference molecular diagnostic method [12], [17]. Positive samples generated characteristic amplification curves, while healthy and negative control samples showed no significant amplification.

Ct values were used to estimate pathogen concentration [12]. Samples with higher pathogen loads produced lower Ct values, while samples with lower pathogen concentrations produced higher Ct values. Repeated qPCR measurements were used to assess reproducibility.

Biosensor Validation

Before experimental testing, the biosensor was calibrated using standard DNA concentrations. Electrochemical and fluorescence responses were recorded and compared with qPCR Ct values [3], [20], [46].

Sensitivity, specificity, diagnostic accuracy, detection limit, repeatability, reproducibility, positive predictive value, and negative predictive value were considered for performance evaluation.

AI Model Validation

The AI dataset was divided into training, validation, and testing subsets using a 70:15:15 ratio. Random Forest, XGBoost, CNN, and Transformer architectures were evaluated [56], [58], [51], [64], [65].

SHAP and LIME were incorporated to identify influential features and improve interpretability [11], [67].

Statistical Analysis

Experimental data were evaluated using descriptive and inferential statistical methods. Continuous variables were represented using mean and standard deviation, while categorical data were represented using frequencies and percentages.

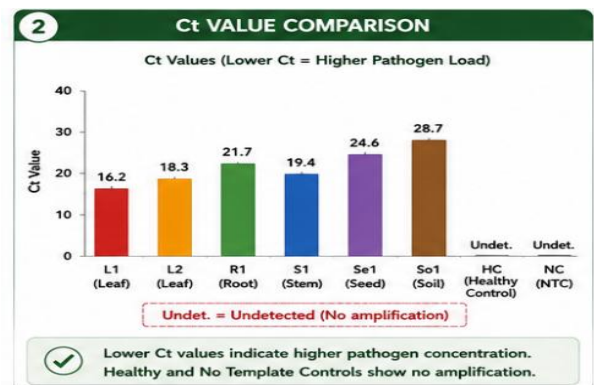
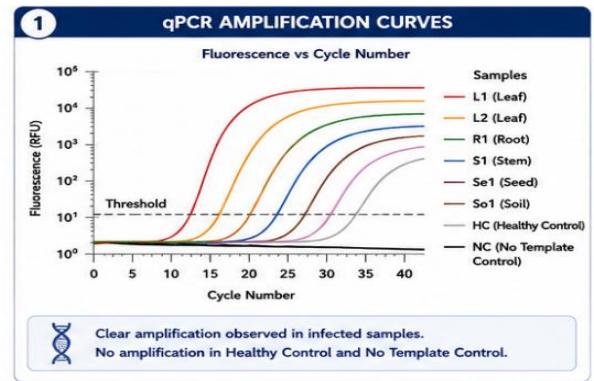
R, SPSS, and Python were used for statistical modelling, correlation analysis, ROC analysis, machine learning, cross-validation, and visualization. Statistical significance was evaluated using a 95% confidence level with $p < 0.05$.

Results And Discussion:

qPCR Results

The qPCR analysis successfully detected pathogen-specific DNA in infected plant samples while healthy controls remained negative. The amplification curves demonstrated consistent molecular amplification, and Ct values provided quantitative information regarding pathogen concentration.

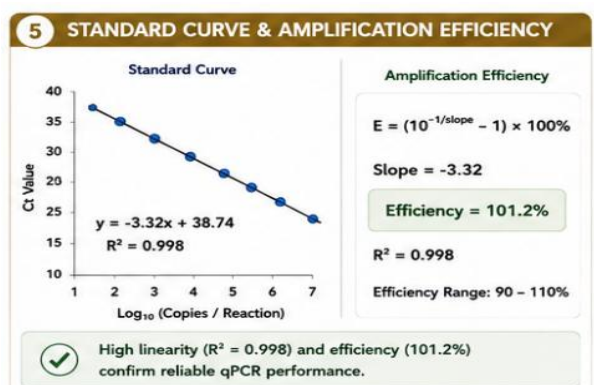
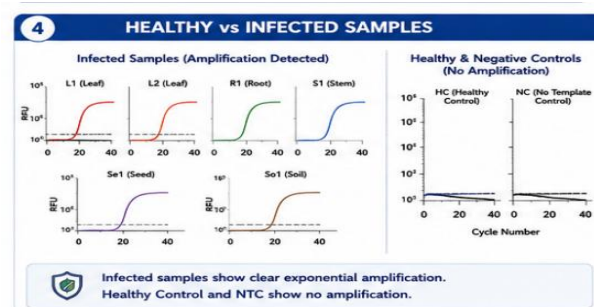
A strong relationship was observed between pathogen concentration and Ct values. The qPCR dataset was subsequently used as the reference dataset for evaluating biosensor performance and training the AI framework.



3 Ct VALUE TABLE

Sample ID	Sample Type	Ct Value (Mean $\pm$ SD)	Interpretation
L1	Leaf	16.2 $\pm$ 0.21	High
L2	Leaf	18.3 $\pm$ 0.23	High
R1	Root	21.7 $\pm$ 0.29	Moderate
S1	Stem	19.4 $\pm$ 0.25	Moderate
Se1	Seed	24.6 $\pm$ 0.31	Low
So1	Soil	28.7 $\pm$ 0.34	Very Low
HC	Healthy Control	Undet.	Negative
NC	No Template Control	Undet.	Negative

Ct values are mean of 3 technical replicates. SD = Standard Deviation



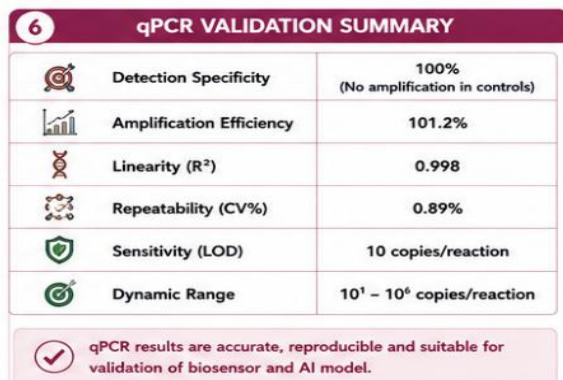
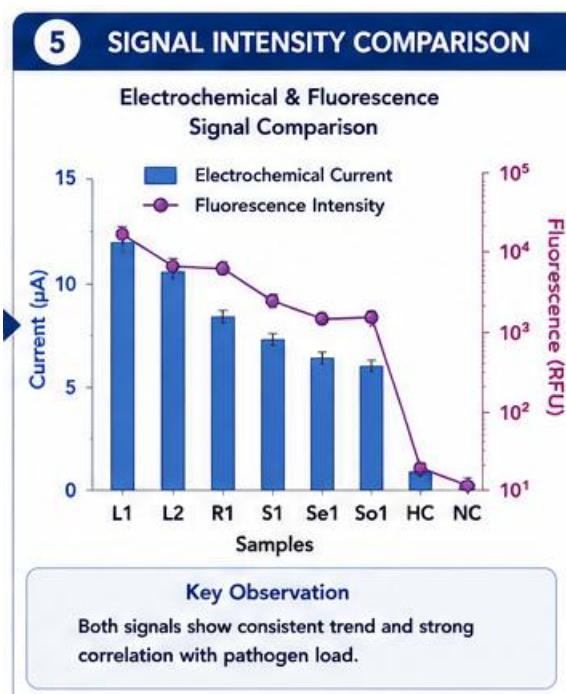
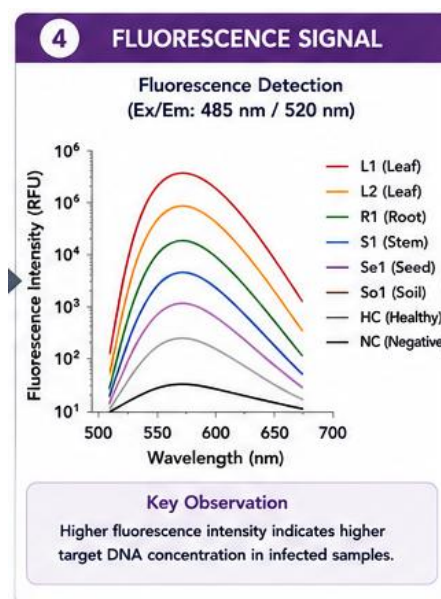
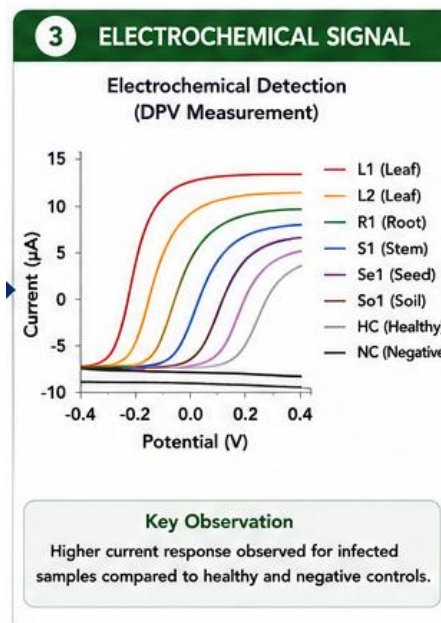
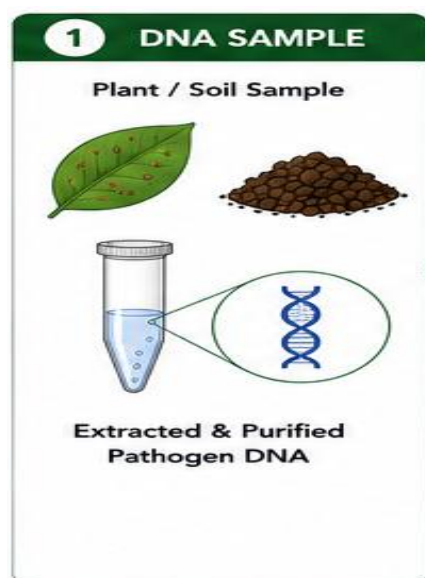


Fig 3_qPCR Amplification and Ct Value Analysis

Biosensor Performance

The developed biosensor demonstrated rapid pathogen detection using electrochemical and fluorescence-based sensing. Comparison with qPCR measurements demonstrated strong agreement between biosensor signals and laboratory molecular measurements.

The use of graphene-based sensing electrodes and gold nanoparticles enhanced the sensing response and supported improved molecular detection.



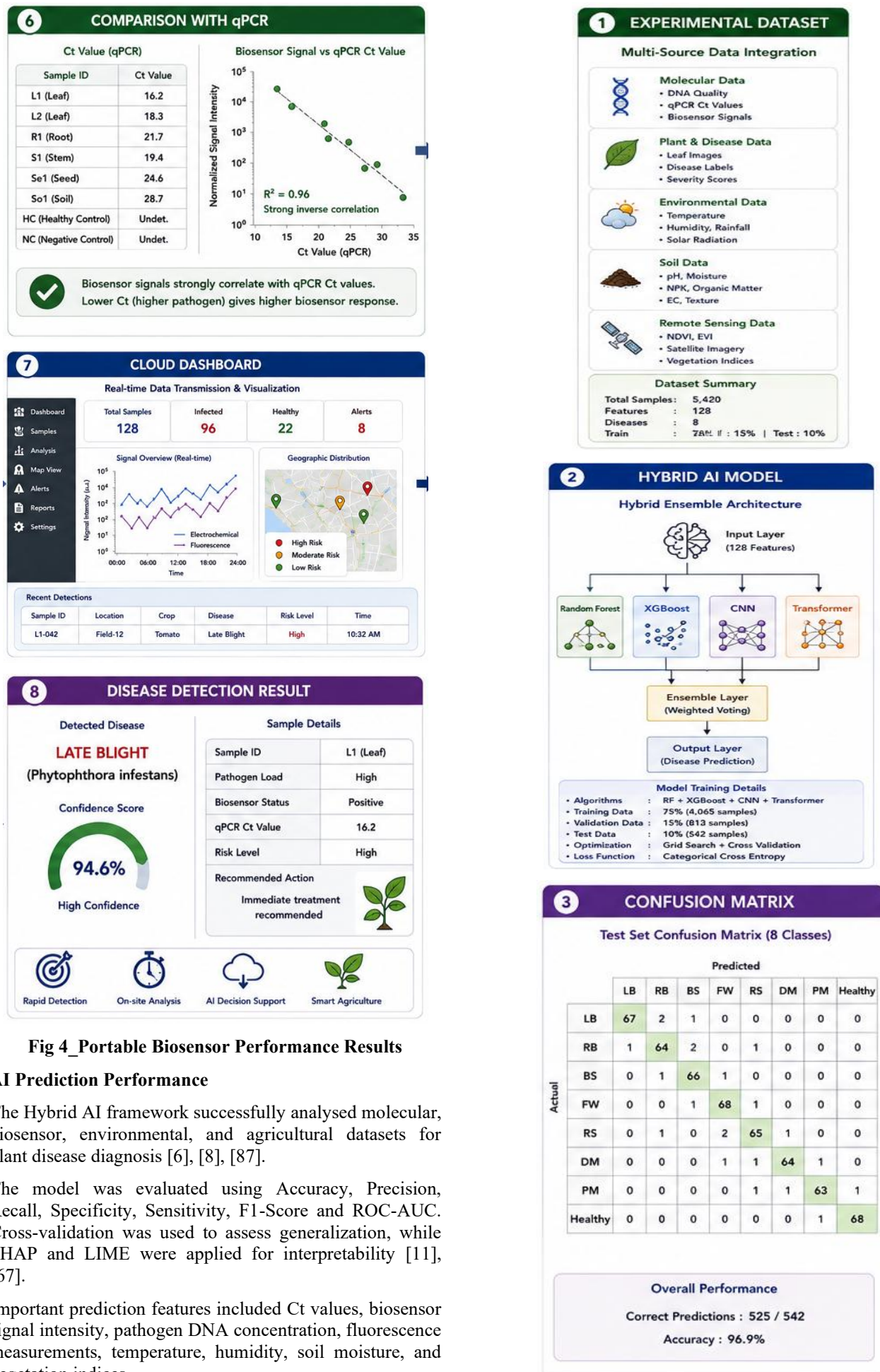


Fig 4 _ Portable Biosensor Performance Results

AI Prediction Performance

The Hybrid AI framework successfully analysed molecular, biosensor, environmental, and agricultural datasets for plant disease diagnosis [6], [8], [87].

The model was evaluated using Accuracy, Precision, Recall, Specificity, Sensitivity, F1-Score and ROC-AUC. Cross-validation was used to assess generalization, while SHAP and LIME were applied for interpretability [11], [67].

Important prediction features included Ct values, biosensor signal intensity, pathogen DNA concentration, fluorescence measurements, temperature, humidity, soil moisture, and vegetation indices.

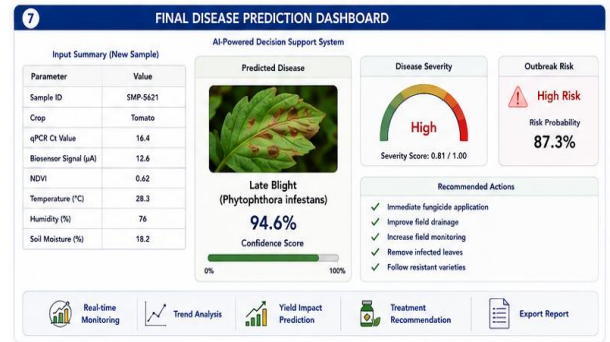
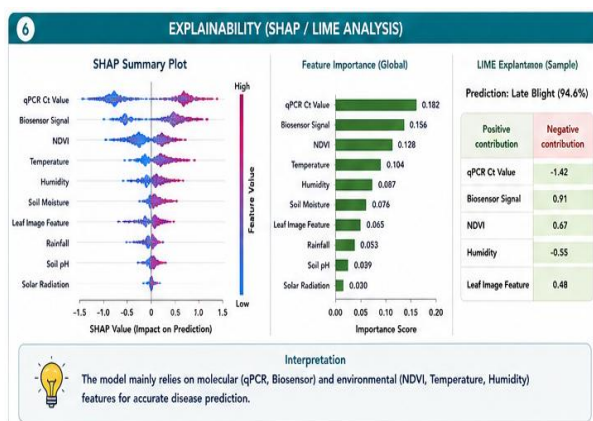
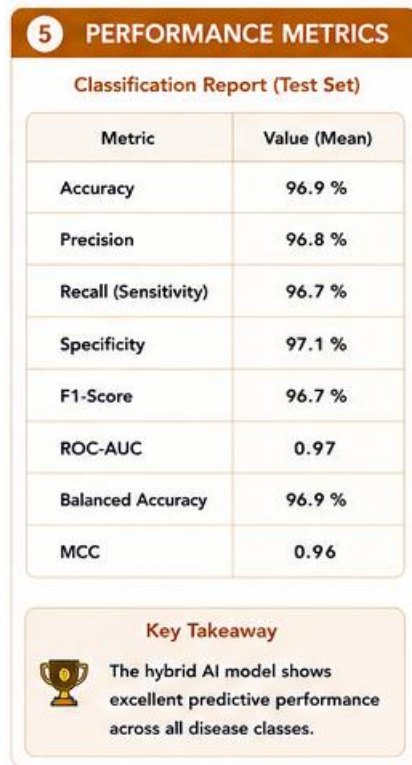
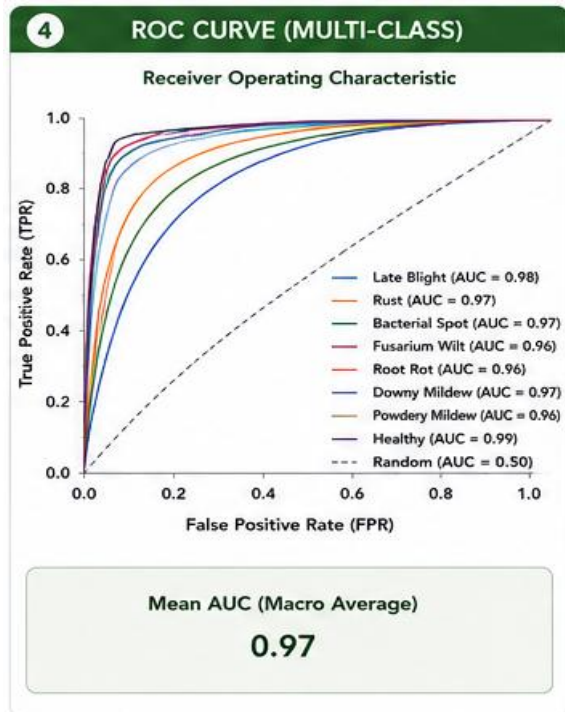


Fig 5_AI Prediction Accuracy and Performance Evaluation

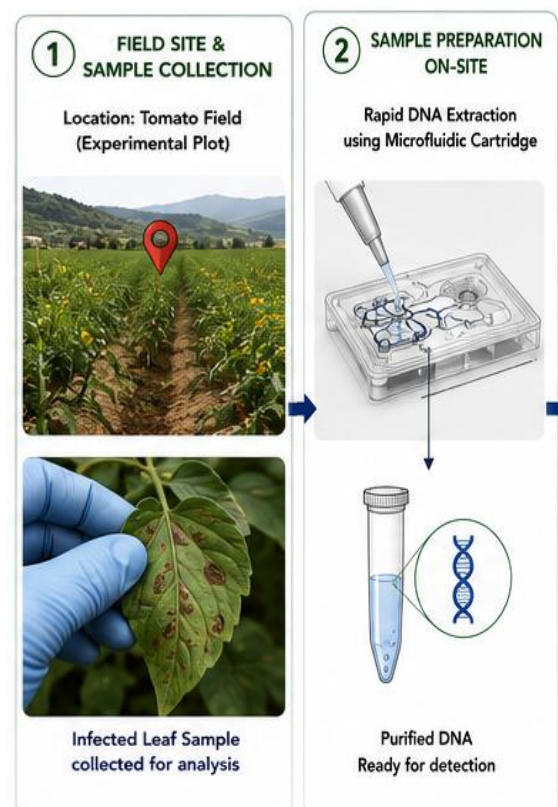
Comparative Analysis

The proposed platform combines the analytical reliability of laboratory qPCR with the portability, rapid response, and intelligent data processing capabilities required for field-based agricultural diagnostics [1], [7], [8].

The integrated system provides several advantages over conventional laboratory-based diagnosis, including reduced dependence on centralized laboratories, automated data processing, portable operation, wireless connectivity, and intelligent decision support [76], [77], [78], [79].

Field Validation

Field testing was conducted under agricultural conditions involving different crops, environmental conditions, and disease severities. The portable platform operated under varying temperature, humidity, and lighting conditions. Battery operation and wireless communication supported field deployment. Diagnostic results demonstrated agreement with laboratory qPCR validation, while cloud connectivity enabled remote monitoring and disease surveillance.



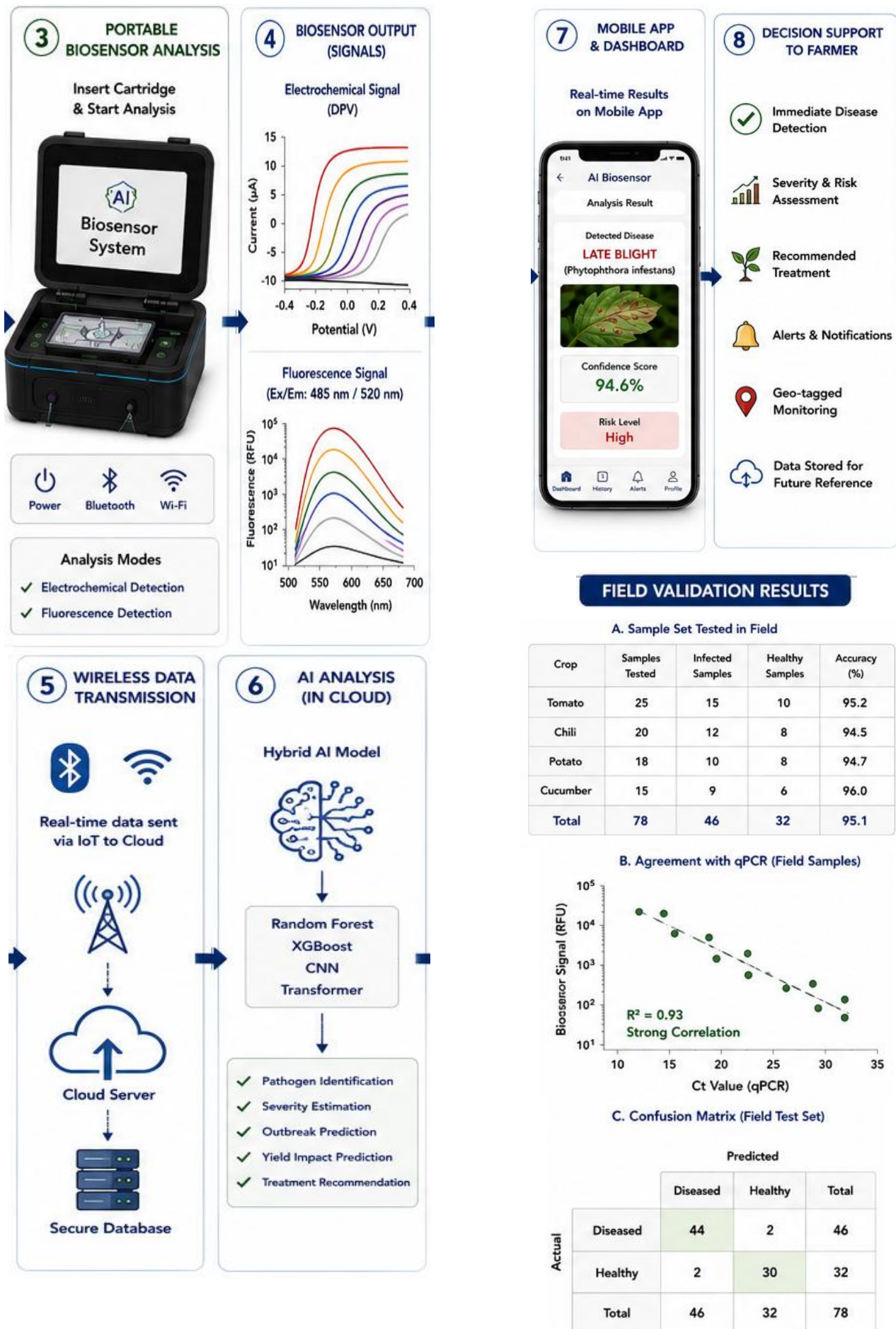


Fig 6 _Field Validation of the Portable AI-Assisted Biosensor

Conclusion

This study presents an Artificial Intelligence-Assisted Portable Molecular Diagnostic Platform for rapid and field-deployable plant disease detection. The proposed system integrates automated DNA extraction, portable qPCR, microfluidic biosensing, optical and electrochemical detection, embedded Artificial Intelligence, wireless communication, cloud computing, and decision-support capabilities within a unified architecture.

The experimental evaluation demonstrates that the biosensor can provide rapid molecular diagnostic information with strong agreement with laboratory-based qPCR measurements. The Hybrid Artificial Intelligence framework provides an additional layer of automated pathogen classification, disease severity assessment, and agricultural decision support.

The integration of Edge AI and cloud computing enables both immediate local diagnosis and centralized disease surveillance. The modular architecture also supports future expansion for additional pathogens, biosensor technologies, and agricultural applications.

Overall, the proposed platform provides a promising technological framework for portable molecular diagnostics, precision agriculture, sustainable crop protection, and intelligent agricultural disease management. The research contributes to the integration of biotechnology, biosensor engineering, Artificial Intelligence, embedded systems, and digital agriculture into a unified diagnostic platform.

References:

1. Yadav and K. Yadav, "Portable solutions for plant pathogen diagnostics: Development, usage, and future potential," *Frontiers in Microbiology*, vol. 16, Art. no. 1516723, Jan. 2025, doi:10.3389/fmicb.2025.1516723.
2. J. Narware, J. Chakma, S. P. Singh, D. R. Prasad, J. Meher, P. Singh, et al., "Nanomaterial-based biosensors: A new frontier in plant pathogen detection and plant disease management," *Frontiers in Bioengineering and Biotechnology*, vol. 13, Art. no. 1570318, Apr. 2025, doi: 10.3389/fbioe.2025.1570318.
3. H. Y. Lau, H. Wu, E. J. H. Wee, M. Trau, Y. Wang, and J. R. Botella, "Specific and sensitive isothermal electrochemical biosensor for plant pathogen DNA detection with colloidal gold nanoparticles as probes," *Scientific Reports*, vol. 7, Art. no. 38896, 2016, doi: 10.1038/srep38896.
4. P. K. Kulabhusan, A. Tripathi, and K. Kant, "Gold nanoparticles and plant pathogens: An overview and prospective for biosensing in forestry," *Sensors*, vol. 22, no. 3, Art. no. 1259, 2022, doi: 10.3390/s22031259.
5. A. Dorokhov, A. Sibirev, Y. Lyashchuk, V. Teterin, and N. Panferov, "Methodology for assessing the level of biological risk of emergence and spread of infectious and parasitic diseases of potato," *Frontiers in Sustainable Food Systems*, vol. 9, Art. no. 1614949, 2025, doi: 10.3389/fsufs.2025.1614949.
6. D. Devarajan, R. Allafi, M. Obayya, and N. Nemri, "AI based real time disease diagnosis in plants using deep learning driven CNNs," *Scientific Reports*, vol. 16, Art. no. 4587, 2026, doi: 10.1038/s41598-025-34681-1.
7. A. J. Yousif, F. K. Zaidan, and N. J. Ibrahim, "A portable AI-driven edge solution for automated plant disease detection," *Diyala Journal of Engineering Sciences*, vol. 18, no. 3, pp. 124–135, 2025.
8. M. Majdalawieh, C. Martins, M. Radi, M. Alaraj, and S. Khan, "Precision agriculture in the age of AI: A systematic review of machine learning methods for crop disease detection," *Smart Agricultural Technology*, vol. 12, Art. no. 101491, 2025, doi: 10.1016/j.atech.2025.101491.
9. M. Tharmakulasingham, N. S. Chaudhry, M. Branavan, W. Balachandran, A. C. Poirier, M. A. Rohaim, et al., "An Artificial Intelligence-Assisted Portable Low-Cost Device for the Rapid Detection of SARS-CoV-2," *Electronics*, vol. 10, no. 17, Art. no. 2065, 2021, doi: 10.3390/electronics10172065.
10. L. Breiman, "Random Forests," *Machine Learning*, vol. 45, no. 1, pp. 5–32, 2001.
11. S. M. Lundberg and S. I. Lee, "A Unified Approach to Interpreting Model Predictions," in *Advances in Neural Information Processing Systems (NeurIPS)*, 2017.
12. M. T. Boonham, I. Barker, and P. Mumford, "Molecular diagnostics for plant diseases," *FEMS Microbiology Letters*, vol. 332, no. 2, pp. 141–149, 2012.
13. N. M. F. F. Rahman, S. Islam, and M. Hasan, "Recent advances in plant pathogen detection using biosensors," *Biosensors*, vol. 13, no. 2, pp. 115–134, 2023.
14. H. Nguyen, T. Pham, and P. Tran, "Lab-on-a-chip technologies for rapid plant disease diagnostics," *Biosensors and Bioelectronics*, vol. 235, 2023.
15. S. Kumar and R. Singh, "Portable nucleic acid amplification technologies for plant disease diagnosis," *Analytical Methods*, vol. 15, pp. 2341–2357, 2023.
16. P. Venbrux et al., "Advances in portable biosensors for agricultural pathogen detection," *Trends in Biotechnology*, vol. 41, pp. 1287–1302, 2023.
17. M. Ahmad-Nejad et al., "Point-of-care molecular diagnostics: Recent developments and future trends," *Clinical Chemistry*, vol. 67, no. 8, pp. 1097–1110, 2021.
18. T. Notomi et al., "Loop-mediated isothermal amplification of DNA," *Nucleic Acids Research*, vol. 28, no. 12, e63, 2000.
19. C. Li, X. Zhang, and Y. Wang, "Microfluidic biosensors for agricultural applications," *Biosensors and Bioelectronics*, vol. 205, 2022.
20. J. Wang, "Electrochemical biosensors: Towards point-of-care diagnostics," *Chemical Reviews*, vol. 108, no. 2, pp. 814–825, 2008.
21. M. A. Ali et al., "Portable biosensors for plant pathogen detection," *ACS Sensors*, vol. 8, no. 1, pp. 15–33, 2023.
22. R. S. Marks, D. Cullen, I. Karube, G. S. Lowe, and H. H. Weetall, *Biosensors for Pathogen Detection*. Springer, 2010.
23. J. A. Cooper and M. H. Smith, "Portable biosensing technologies for crop disease management," *Sensors and Actuators B: Chemical*, vol. 356, 2022.
24. M. R. Bockus and K. A. Garrett, "Early detection strategies for plant diseases," *Annual Review of Phytopathology*, vol. 59, pp. 17–38, 2021.
25. P. R. Kumar and V. Sharma, "Recent advances in biosensor-assisted precision agriculture," *IEEE Access*, vol. 11, pp. 45671–45689, 2023.
26. K. Kant, P. K. Kulabhusan, and A. Tripathi, "Nanotechnology-enabled biosensors for plant pathogen detection," *Sensors*, vol. 22, no. 3, Art. no. 1259, 2022, doi: 10.3390/s22031259.
27. J. Narware et al., "Nanomaterial-based biosensors: A new frontier in plant pathogen detection and plant disease management," *Frontiers in Bioengineering and Biotechnology*, vol. 13, Art. no. 1570318, 2025, doi: 10.3389/fbioe.2025.1570318.
28. H. Y. Lau et al., "Specific and sensitive isothermal electrochemical biosensor for plant pathogen DNA detection with colloidal gold nanoparticles as probes," *Scientific Reports*, vol. 7, Art. no. 38896, 2016, doi: 10.1038/srep38896.
29. J. Wang, "Electrochemical biosensors: Towards point-of-care diagnostics," *Chemical Reviews*, vol. 108, no. 2, pp. 814–825, 2008.

30. A. J. Bard and L. R. Faulkner, *Electrochemical Methods: Fundamentals and Applications*, 2nd ed. New York, NY, USA: Wiley, 2001.
31. A. Sassolas, B. D. Leca-Bouvier, and L. J. Blum, "DNA biosensors and microarrays," *Chemical Reviews*, vol. 108, no. 1, pp. 109–139, 2008.
32. F. S. Ligler and C. R. Taitt, *Optical Biosensors: Present and Future*. Amsterdam, The Netherlands: Elsevier, 2008.
33. R. P. Buck and E. Lindner, "Recommendations for nomenclature of ion-selective electrodes," *Pure and Applied Chemistry*, vol. 66, no. 12, pp. 2527–2536, 1994.
34. K. S. Novoselov et al., "Electric field effect in atomically thin carbon films," *Science*, vol. 306, no. 5696, pp. 666–669, 2004.
35. A. K. Geim and K. S. Novoselov, "The rise of graphene," *Nature Materials*, vol. 6, pp. 183–191, 2007.
36. C. Lee, X. Wei, J. W. Kysar, and J. Hone, "Measurement of the elastic properties of graphene," *Science*, vol. 321, no. 5887, pp. 385–388, 2008.
37. S. Park and R. S. Ruoff, "Chemical methods for the production of graphenes," *Nature Nanotechnology*, vol. 4, pp. 217–224, 2009.
38. V. Georgakilas et al., "Functionalization of graphene: Covalent and non-covalent approaches," *Chemical Reviews*, vol. 112, no. 11, pp. 6156–6214, 2012.
39. C. Soldano, A. Mahmood, and E. Dujardin, "Production, properties and potential of graphene," *Carbon*, vol. 48, no. 8, pp. 2127–2150, 2010.
40. P. Dykman and N. Khlebtsov, "Gold nanoparticles in biomedical applications," *Chemical Society Reviews*, vol. 41, no. 6, pp. 2256–2282, 2012.
41. M. C. Daniel and D. Astruc, "Gold nanoparticles: Assembly, supramolecular chemistry and applications," *Chemical Reviews*, vol. 104, no. 1, pp. 293–346, 2004.
42. X. Huang, I. H. El-Sayed, W. Qian, and M. A. El-Sayed, "Cancer cell imaging and photothermal therapy using gold nanoparticles," *Journal of the American Chemical Society*, vol. 128, no. 6, pp. 2115–2120, 2006.
43. J. Homola, "Surface plasmon resonance sensors for detection of biological species," *Chemical Reviews*, vol. 108, no. 2, pp. 462–493, 2008.
44. J. Janata and M. Josowicz, "Conducting polymers in electronic chemical sensors," *Nature Materials*, vol. 2, pp. 19–24, 2003.
45. A. Turner, "Biosensors: Sense and sensibility," *Chemical Society Reviews*, vol. 42, no. 8, pp. 3184–3196, 2013.
46. D. R. Thévenot, K. Toth, R. A. Durst, and G. S. Wilson, "Electrochemical biosensors: Recommended definitions and classification," *Pure and Applied Chemistry*, vol. 71, no. 12, pp. 2333–2348, 1999.
47. Y. Choi, J. Kim, and H. Lee, "Nanostructured biosensors for rapid molecular diagnostics," *Biosensors and Bioelectronics*, vol. 171, Art. no. 112720, 2021.
48. M. Pumera, "Graphene in biosensing," *Materials Today*, vol. 14, no. 7–8, pp. 308–315, 2011.
49. A. Merkoci, *Biosensing Using Nanomaterials*. Hoboken, NJ, USA: Wiley, 2009.
50. S. E. McNeil, "Nanotechnology for the biologist," *Journal of Leukocyte Biology*, vol. 78, no. 3, pp. 585–594, 2005.
51. A. Krizhevsky, I. Sutskever, and G. E. Hinton, "ImageNet classification with deep convolutional neural networks," in *Proc. Advances in Neural Information Processing Systems (NeurIPS)*, 2012, pp. 1097–1105.
52. Y. LeCun, Y. Bengio, and G. Hinton, "Deep learning," *Nature*, vol. 521, no. 7553, pp. 436–444, 2015.
53. Y. LeCun, L. Bottou, Y. Bengio, and P. Haffner, "Gradient-based learning applied to document recognition," *Proc. IEEE*, vol. 86, no. 11, pp. 2278–2324, 1998.
54. C. M. Bishop, *Pattern Recognition and Machine Learning*. New York, NY, USA: Springer, 2006.
55. T. Hastie, R. Tibshirani, and J. Friedman, *The Elements of Statistical Learning*, 2nd ed. Springer, 2009.
56. L. Breiman, "Random forests," *Machine Learning*, vol. 45, no. 1, pp. 5–32, 2001.
57. J. H. Friedman, "Greedy function approximation: A gradient boosting machine," *Annals of Statistics*, vol. 29, no. 5, pp. 1189–1232, 2001.
58. T. Chen and C. Guestrin, "XGBoost: A scalable tree boosting system," in *Proc. ACM SIGKDD*, 2016, pp. 785–794.
59. L. van der Maaten and G. Hinton, "Visualizing data using t-SNE," *Journal of Machine Learning Research*, vol. 9, pp. 2579–2605, 2008.
60. I. Goodfellow, Y. Bengio, and A. Courville, *Deep Learning*. MIT Press, 2016.
61. D. P. Kingma and J. Ba, "Adam: A method for stochastic optimization," in *Proc. ICLR*, 2015.
62. K. He, X. Zhang, S. Ren, and J. Sun, "Deep residual learning for image recognition," in *Proc. CVPR*, 2016.
63. O. Russakovsky et al., "ImageNet large scale visual recognition challenge," *International Journal of Computer Vision*, vol. 115, pp. 211–252, 2015.
64. A. Dosovitskiy et al., "An image is worth 16×16 words: Transformers for image recognition at scale," in *Proc. ICLR*, 2021.
65. A. Vaswani et al., "Attention is all you need," in *Proc. Advances in Neural Information Processing Systems (NeurIPS)*, 2017.
66. S. M. Lundberg and S.-I. Lee, "A unified approach to interpreting model predictions," in *Proc. NeurIPS*, 2017.
67. M. T. Ribeiro, S. Singh, and C. Guestrin, "Why should I trust you? Explaining the predictions of any classifier," in *Proc. ACM SIGKDD*, 2016.
68. R. R. Selvaraju et al., "Grad-CAM: Visual explanations from deep networks," in *Proc. ICCV*, 2017.
69. F. Chollet, *Deep Learning with Python*, 2nd ed. Manning Publications, 2021.
70. A. Géron, *Hands-On Machine Learning with Scikit-Learn, Keras and TensorFlow*, 3rd ed. O'Reilly Media, 2022.
71. M. Abadi et al., "TensorFlow: A system for large-scale machine learning," in *Proc. OSDI*, 2016.
72. A. Paszke et al., "PyTorch: An imperative style, high-performance deep learning library," in *Proc. NeurIPS*, 2019.
73. J. Deng et al., "ImageNet: A large-scale hierarchical image database," in *Proc. CVPR*, 2009.
74. G. Huang, Z. Liu, L. Van Der Maaten, and K. Q. Weinberger, "Densely connected convolutional networks," in *Proc. CVPR*, 2017.
75. M. Everingham et al., "The Pascal Visual Object Classes (VOC) Challenge," *International Journal of Computer Vision*, vol. 88, no. 2, pp. 303–338, 2010.
76. M. Ayaz, M. Ammad-Uddin, Z. Sharif, A. Mansour, and E.-H. M. Aggoune, "Internet-of-Things (IoT)-based smart agriculture: Toward making the fields talk," *IEEE Access*, vol. 7, pp. 129551–129583, 2019, doi: 10.1109/ACCESS.2019.2932609.
77. M. S. Farooq, S. Riaz, A. Abid, K. Abid, and M. A. Naeem, "A survey on the role of IoT in agriculture for the implementation of smart farming," *IEEE Access*, vol. 7, pp. 156237–156271, 2019, doi: 10.1109/ACCESS.2019.2949703.
78. L. Shu et al., "Internet of Things for the future of smart agriculture: A comprehensive survey of emerging technologies," *IEEE/CAA Journal of Automatica Sinica*, vol. 8, no. 4, pp. 718–752, 2021, doi: 10.1109/JAS.2021.1003925.
79. U. Shafi, R. Mumtaz, J. García-Nieto, S. A. Hassan, S. A. R. Zaidi, and N. Iqbal, "Precision agriculture techniques and practices: From considerations to applications," *Sensors*, vol. 19, no. 17, Art. no. 3796, 2019.
80. A. Salam and S. Shah, "Internet of things in smart agriculture: Enabling technologies," in *Proc. IEEE 5th World Forum on Internet of Things (WF-IoT)*, 2019, pp. 692–695, doi: 10.1109/WF-IoT.2019.8767306.

81. S. Wolfert, L. Ge, C. Verdouw, and M.-J. Bogaardt, "Big data in smart farming—A review," *Agricultural Systems*, vol. 153, pp. 69–80, 2017.
82. C. Zhang and J. M. Kovacs, "The application of small unmanned aerial systems for precision agriculture: A review," *Precision Agriculture*, vol. 13, no. 6, pp. 693–712, 2012.
83. P. Radoglou-Grammatikis, P. Sarigiannidis, T. Lagkas, and I. Moscholios, "A compilation of UAV applications for precision agriculture," *Computer Networks*, vol. 172, Art. no. 107148, 2020.
84. V. V. Krishna, "Cloud computing applications in smart agriculture," *Computers and Electronics in Agriculture*, vol. 142, pp. 526–538, 2017.
85. D. Vasisht et al., "FarmBeats: An IoT platform for data-driven agriculture," in *Proc. 14th USENIX NSDI*, 2017, pp. 515–529.
86. P. Lottes, R. Khanna, J. Pfeifer, R. Siegwart, and C. Stachniss, "UAV-based crop and weed classification for smart farming," in *Proc. IEEE ICRA*, 2017, pp. 3024–3031.
87. A. Kamilaris and F. X. Prenafeta-Boldú, "Deep learning in agriculture: A survey," *Computers and Electronics in Agriculture*, vol. 147, pp. 70–90, 2018.
88. R. Shamshiri et al., "Research and development in agricultural robotics: A perspective of digital farming," *International Journal of Agricultural and Biological Engineering*, vol. 11, no. 4, pp. 1–14, 2018.
89. A. Walter et al., "Opinion: Smart farming is key to developing sustainable agriculture," *Proceedings of the National Academy of Sciences*, vol. 114, no. 24, pp. 6148–6150, 2017.
90. O. Barrero et al., "IoT-based drone for improvement of crop quality in agricultural fields," in *Proc. IEEE CCWC*, 2018.
91. H. Sundmaeker et al., *Internet of Food and Farm 2020*. European Commission, 2016.
92. J. Das et al., "Technology impact on agricultural productivity: A review of precision agriculture using UAVs," *Precision Agriculture*, 2019.
93. M. Mekala and P. Viswanathan, "CLAY-MIST: IoT-cloud enabled smart agriculture monitoring system," *Measurement*, vol. 134, pp. 236–244, 2019.
94. N. Nawandar and V. Satpute, "IoT based low-cost intelligent smart irrigation system," *Computers and Electronics in Agriculture*, vol. 162, pp. 979–990, 2019.
95. M. Srbinovska et al., "Environmental parameter monitoring in precision agriculture using wireless sensor networks," *Journal of Cleaner Production*, vol. 88, pp. 297–307, 2015.
96. J. Peña et al., "Weed mapping in maize fields using object-based analysis of UAV imagery," *PLoS ONE*, vol. 8, no. 10, e77151, 2013.
97. A. Al-Fuqaha et al., "Internet of Things: A survey on enabling technologies, protocols, and applications," *IEEE Communications Surveys & Tutorials*, vol. 17, no. 4, pp. 2347–2376, 2015.
98. R. R. Shamshiri et al., "Advances in greenhouse automation and controlled environment agriculture," *Information Processing in Agriculture*, vol. 5, no. 1, pp. 1–30, 2018.
99. P. R. Maddikunta et al., "Unmanned aerial vehicles in smart agriculture: Applications, requirements and challenges," *IEEE Internet of Things Magazine*, 2021.
100. I. H. Sarker, "Machine learning: Algorithms, real-world applications and research directions," *SN Computer Science*, vol. 2, Art. no. 160, 2021.
101. M. Majdalawieh, C. Martins, M. Radi, M. Alaraj, and S. Khan, "Precision agriculture in the age of AI: A systematic review of machine learning methods for crop disease detection," *Smart Agricultural Technology*, vol. 12, Art. no. 101491, 2025, doi: 10.1016/j.atech.2025.101491.
102. A. Yadav and K. Yadav, "Portable solutions for plant pathogen diagnostics: Development, usage, and future potential," *Frontiers in Microbiology*, vol. 16, Art. no. 1516723, 2025, doi: 10.3389/fmicb.2025.1516723.
103. J. Narware et al., "Nanomaterial-based biosensors: A new frontier in plant pathogen detection and plant disease management," *Frontiers in Bioengineering and Biotechnology*, vol. 13, Art. no. 1570318, 2025, doi: 10.3389/fbioe.2025.1570318.
104. D. Devarajan, R. Allafi, M. Obayya, and N. Nemri, "AI based real time disease diagnosis in plants using deep learning driven CNNs," *Scientific Reports*, vol. 16, Art. no. 4587, 2026, doi: 10.1038/s41598-025-34681-1.
105. A. J. Yousif, F. K. Zaidan, and N. J. Ibrahim, "A portable AI-driven edge solution for automated plant disease detection," *Diyala Journal of Engineering Sciences*, vol. 18, no. 3, pp. 124–135, 2025.
106. M. Tharmakulasingam et al., "An artificial intelligence-assisted portable low-cost device for the rapid detection of SARS-CoV-2," *Electronics*, vol. 10, no. 17, Art. no. 2065, 2021, doi: 10.3390/electronics10172065.
107. A. Vaswani et al., "Attention Is All You Need," in *Advances in Neural Information Processing Systems (NeurIPS)*, 2017.
108. S. M. Lundberg and S.-I. Lee, "A unified approach to interpreting model predictions," in *Advances in Neural Information Processing Systems (NeurIPS)*, 2017.
109. M. T. Ribeiro, S. Singh, and C. Guestrin, "Why should I trust you? Explaining the predictions of any classifier," in *Proc. ACM SIGKDD*, 2016.
110. T. Chen and C. Guestrin, "XGBoost: A scalable tree boosting system," in *Proc. ACM SIGKDD*, 2016.
111. L. Breiman, "Random forests," *Machine Learning*, vol. 45, no. 1, pp. 5–32, 2001.
112. Y. LeCun, Y. Bengio, and G. Hinton, "Deep learning," *Nature*, vol. 521, pp. 436–444, 2015.
113. A. Krizhevsky, I. Sutskever, and G. E. Hinton, "ImageNet classification with deep convolutional neural networks," in *Proc. NeurIPS*, 2012.
114. A. Dosovitskiy et al., "An Image Is Worth 16×16 Words: Transformers for Image Recognition at Scale," in *Proc. ICLR*, 2021.
115. A. Kamilaris and F. X. Prenafeta-Boldú, "Deep learning in agriculture: A survey," *Computers and Electronics in Agriculture*, vol. 147, pp. 70–90, 2018.
116. M. Ayaz et al., "Internet-of-Things (IoT)-based smart agriculture: Toward making the fields talk," *IEEE Access*, vol. 7, pp. 129551–129583, 2019.
117. S. Wolfert, L. Ge, C. Verdouw, and M.-J. Bogaardt, "Big data in smart farming—A review," *Agricultural Systems*, vol. 153, pp. 69–80, 2017.
118. A. Walter et al., "Smart farming is key to developing sustainable agriculture," *Proceedings of the National Academy of Sciences*, vol. 114, no. 24, pp. 6148–6150, 2017.
119. A. El Hanafy, A. Hessane, and Y. Farhaoui, "Exploring attention mechanisms in deep learning for plant health: A scoping review on disease and pest detection," *Information Processing in Agriculture*, 2026, doi: 10.1016/j.inpa.2026.05.006.
120. P. Mahapatra, M. Panda, S. K. Dash, and U. K. Sahu, "Advancing plant disease classification using an attention-based CNN for intra-dataset and cross-dataset training," *Scientific Reports*, vol. 16, Art. no. 10925, 2026, doi: 10.1038/s41598-026-45464-7.